



H2020-MSCA ITN  
Grant n. 956099



## Optimal data acquisition of biological samples

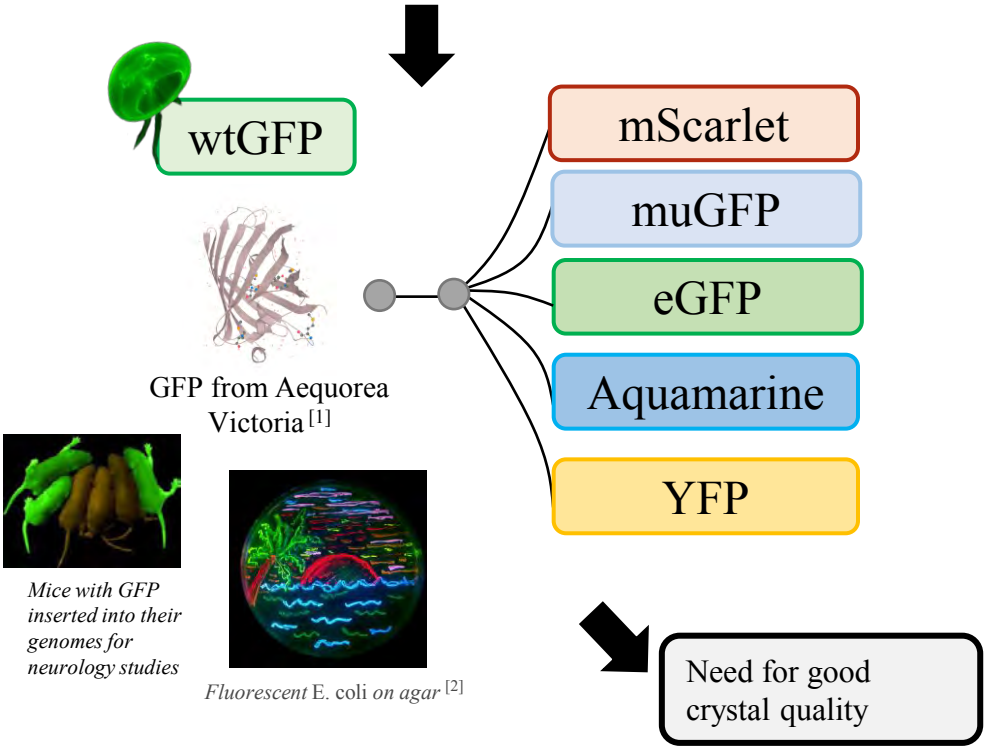
*Boosting data quality of electron diffraction through  
novel technologies*



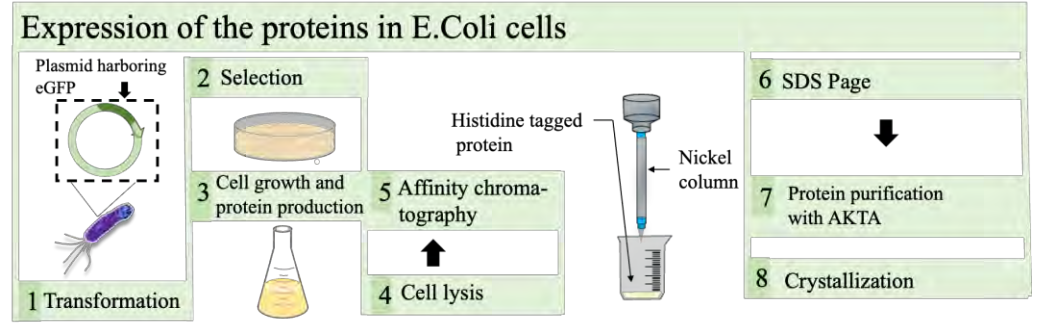
**BIOZENTRUM**

Universität Basel  
The Center for Molecular Life Sciences

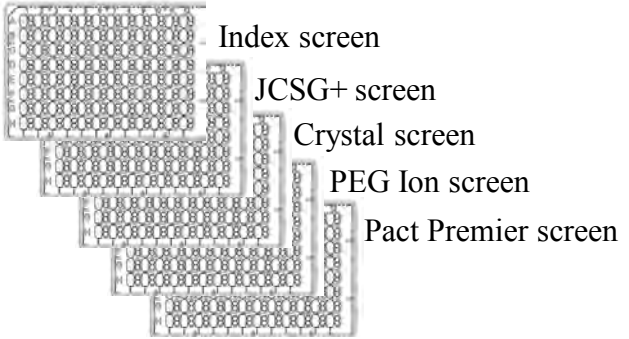
**Structural damage vs functional damage**  
 (Decay of Bragg peaks intensity) ⚡ (fluorescence)



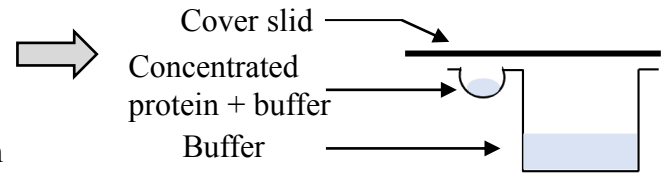
**eGFP crystals preparation:**



**Commercial screening kits**



**Sitting drop vapor diffusion method**



[1] Deposited: Aug 1996, Ormo, M. et al. [2] Fluorescent E. coli on agar, Nathan Shaner, photography by Paul Steinbach, created in the lab of Nobel Prize winner Roger Tsien

Nice rod like eGFP crystals...

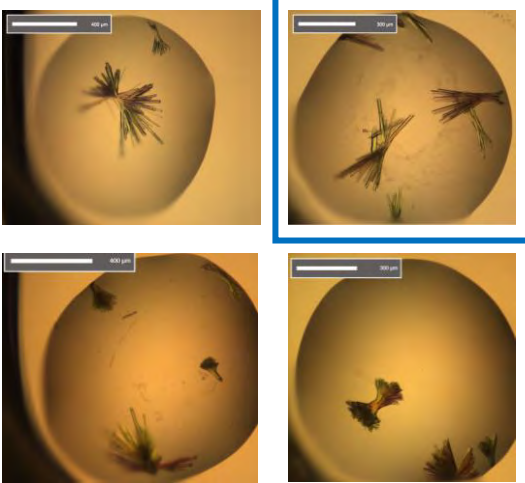
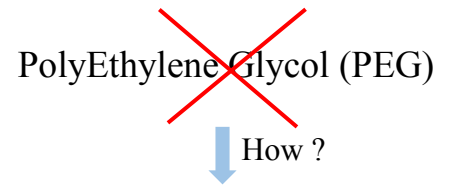


Figure 1: eGFP crystals images taken with the RockImager

...but too big

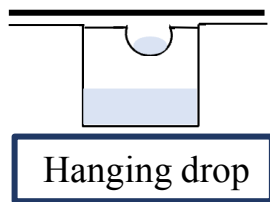
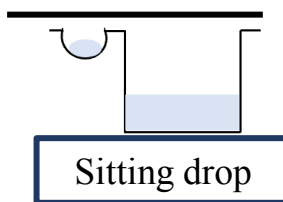
How to reduce their size without breaking or slicing them?

Try to grow crystals with suitable size in the first place. Avoid as much as possible the viscosity of the growth environment to produce better quality grids.



Add salt (NaCl) to a hit with big crystals and reduce PEG concentration.

REF Condition  
[Prot] = 17mg/mL  
25% PEG 1500  
MMT 1M pH5



- Increase the concentration of proteine
- Add salt at different concentration in the droplet only and keep the ref buffer
- Add salt at different concentration in the reservoir and not in the droplet
- Add salt at different concentrations in the droplet and in the reservoir
- Change the ratio protein:buffer in the droplet
- Increase the droplet volume without changing the volume of the reservoir
- .....

« If you can see them with the light microscope, they are already too big »

**Effect of adding salt in the droplet only**  
Well 400 µL Buffer B1  
Fixed eGFP concentration: [19.820 mg/mL]

| Droplet: 1µL protéine + 1 µL B1<br>Well 400 µL B1 | Droplet: 1µL protéine + 1µL e1<br>Well 400 µL B1 | Droplet: 1 µL protéine + 1 µL e2<br>Well 400 µL B1 | Droplet: 1 µL protéine + 1 µL e3<br>Well 400 µL B1 |
|---|--|--|--|
|   |  |  |  |

**Effect of adding salt in the droplet, and change the ratio prot:buffer in the droplet**  
Well 400 µL Buffer B1  
Fixed eGFP concentration: [19.820 mg/mL]

| Droplet: 1.5 µL protéine + 0.5 µL B1<br>Well 400 µL e3 | Droplet: 1.5 µL protéine + 0.5 µL e1<br>Well 400 µL e2 | Droplet: 1.5 µL protéine + 0.5 µL e2<br>Well 400 µL b1 | Droplet: 1.5 µL protéine + 0.5 µL e3<br>Well 400 µL e1 |
|--|--|--|--|
|  |  |  |  |

Before going to the TEM to check for diffraction...

# SONICC

*Second order non-linear  
Imaging of Chiral crystals*

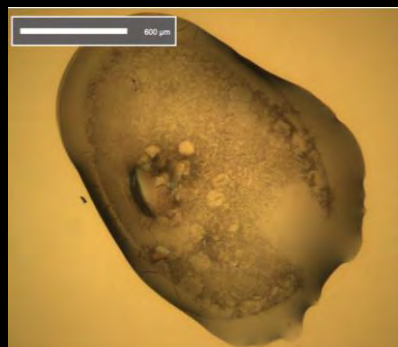
- ❑ To find crystals buried in precipitate or detect microcrystallinity in the drop.
- ❑ Crystals appear white against a stark black background and can therefore be distinguished from amorphous precipitate.



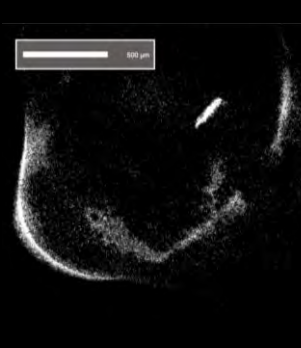
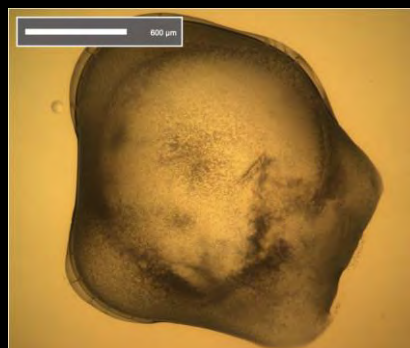
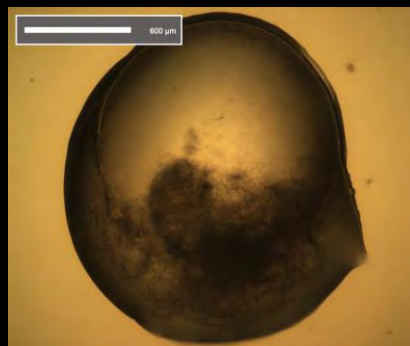
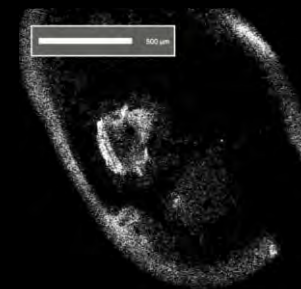
A hit ?

Unsuccessful too icy grids...

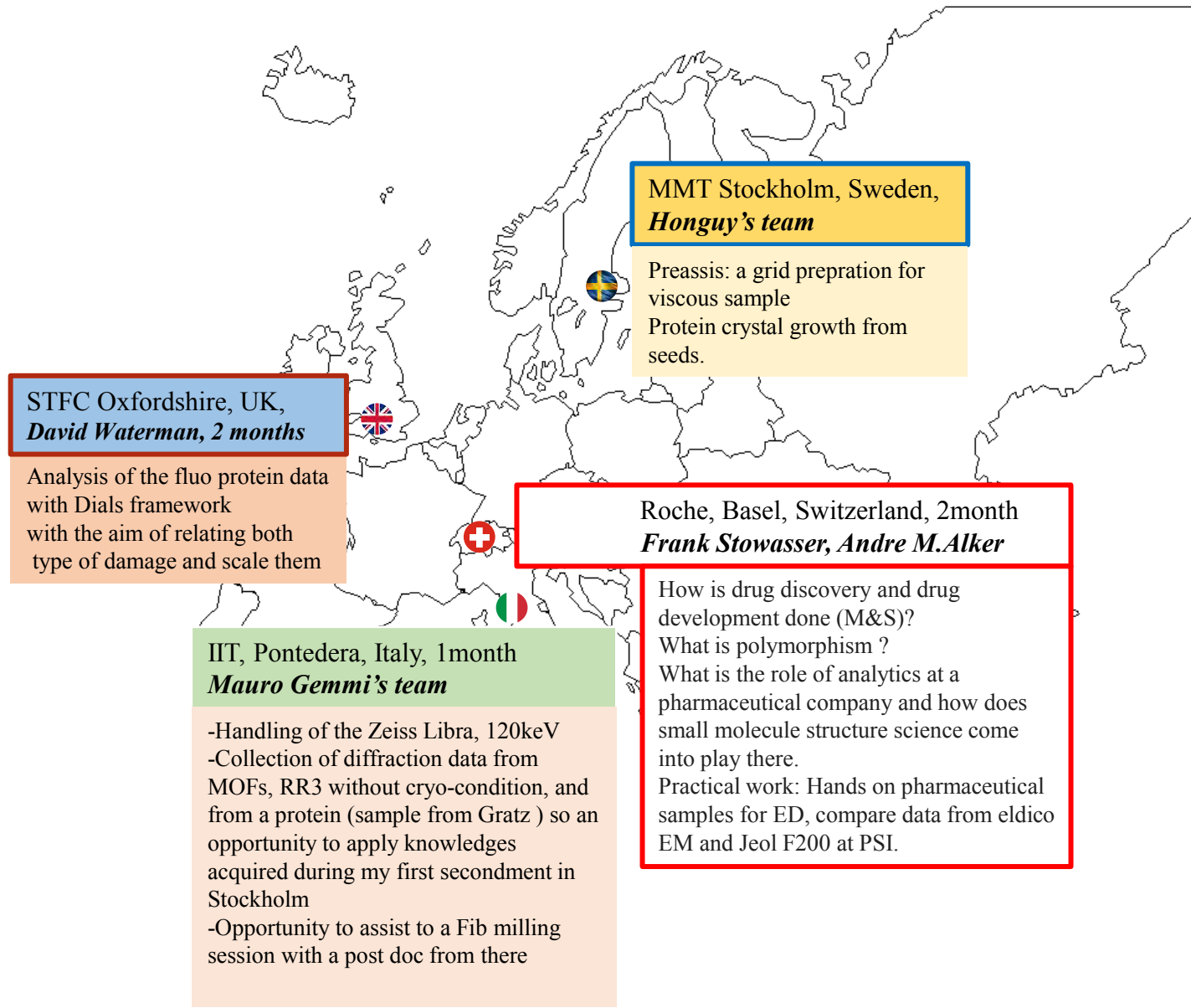
Visible Light



SHG



To be followed





Versailles Conference

MMT Stockholm, Sweden,  
*Honguy's team*



Biozentrum, Basel, PSI, Villigen, Switzerland



IIT, Pontedera, Italy, 1month  
*Mauro Gemmi's team*



**THANK YOU**

